

# Using `asreml` and `asremlPlus` for the Ladybird example from Welham et al. (2014)

Chris Brien

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## Introduction

This vignette shows how to use `asremlPlus` (Brien, 2024a), and `dae` (Brien, 2024b), for exploring and presenting predictions (estimated marginal means: EMMs) from a linear mixed model analysis, the predictions having been produced using `asreml` (Butler et al., 2023). Here, `asremlPlus`, `dae` and `asreml` are packages for the R Statistical Computing environment (R Core Team, 2024).

The context is a three-factor factorial experiment on ladybirds (Welham et al., 2014, Example 8.2) that aims to answer the question “Will ladybirds transfer fungus to aphids on plants?” The experiment consists of 2 runs of 36 containers, each with a plant and aphids. There are three factors that results in 12 treatments: Host plant (beans, trefoil), infected Cadavers (5, 10, 20), Ladybird (-, +). These are randomized to the containers within a run so that each is replicated 3 times within a run. The response to be analysed is the logit of the proportion of live aphids that were infected.

## Initialize

```
library(knitr)
opts_chunk$set("tidy" = FALSE, comment = NA)
suppressMessages(library(asreml))
```

```
## Offline License checked out Tue Dec 10 17:00:54 2024
```

```
packageVersion("asreml")
```

```
## [1] '4.2.0.332'
```

```
suppressMessages(library(asremlPlus))
packageVersion("asremlPlus")
```

```
## [1] '4.4.43'
```

```
suppressMessages(library("dae"))
packageVersion("dae")
```

```
## [1] '3.2.30'
```

```
options(width = 95, show.signif.stars = FALSE)
```

## Get data available in asremlPlus

```
data("Ladybird.dat")
```

## Do an ANOVA of logits

```
Ladybird.aov <- aov(logitP ~ Host*Cadavers*Ladybird + Error(Run/Plant),  
                   data=Ladybird.dat)  
summary(Ladybird.aov)
```

Error: Run

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Residuals	1	0.06766	0.06766		

Error: Run:Plant

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Host	1	13.599	13.599	59.172	1.82e-10
Cadavers	2	17.027	8.514	37.044	3.78e-11
Ladybird	1	11.091	11.091	48.257	3.33e-09
Host:Cadavers	2	0.308	0.154	0.670	0.5158
Host:Ladybird	1	0.228	0.228	0.992	0.3234
Cadavers:Ladybird	2	1.735	0.867	3.774	0.0287
Host:Cadavers:Ladybird	2	0.200	0.100	0.435	0.6493
Residuals	59	13.560	0.230		

The anova table gives the F-tests for the three-factor effects and interactions. Note the **Residuals Mean Sq** value for Run:Plant of 0.230. Also, it is clear that the Run component is negative, given that the **Residuals Mean Sq** value for Run is less than that for Run:Plant; it is  $(0.06766 - 0.230) / 36$ . From the table it is seen that the only significant interaction is Cadavers:Ladybird and that the Host main effect is significant.

## Use asreml to analyse the logits

### Mixed model analysis of logits

```
m <- asreml(logitP ~ Host*Cadavers*Ladybird,  
            random = ~ Run,  
            residual = ~ Run:Plant,  
            data = Ladybird.dat)
```

ASReml Version 4.2 10/12/2024 17:00:54

	LogLik	Sigma2	DF	wall
1	3.071301	0.2262391	60	17:00:54 ( 1 restrained)

```

2      3.642605      0.2269125      60  17:00:54 ( 1 restrained)
3      3.712498      0.2271056      60  17:00:54 ( 1 restrained)
4      3.717213      0.2271206      60  17:00:54 ( 1 restrained)
5      3.717512      0.2271216      60  17:00:54 ( 1 restrained)

```

```
summary(m)$varcomp
```

```

          component  std.error  z.ratio bound %ch
Run          2.298309e-08      NA      NA      B  NA
Run:Plant!R  2.271216e-01  0.04156985  5.463612      P  0

```

As expected the Run component is bound (B) at approximately zero. This results in a change in the estimate of the residual variance to 0.227. To allow for a negative estimate we will unconstrain the Run component. As Littell et al. (2006, p.150) say

if you do not set the negative variance component estimate to zero, but allow it to remain negative, you get better control over Type I error and, for cases of negative wholeplot error variance estimates, greater power. Therefore, this is the recommended procedure.

### Unconstrain Repls to make the analysis equivalent to ANOVA

```
m <- setvarianceterms(m$call, terms = "Run", bounds = "U")
```

```
ASReml Version 4.2 10/12/2024 17:00:55
```

```

          LogLik          Sigma2      DF      wall
1      3.071301      0.2262391      60  17:00:55 ( 1 restrained)
2      3.642605      0.2269125      60  17:00:55 ( 1 restrained)
3      3.802834      0.2274541      60  17:00:55 ( 1 restrained)
4      3.839487      0.2334176      60  17:00:55
5      3.955683      0.2309710      60  17:00:55
6      3.973334      0.2300353      60  17:00:55
7      3.974051      0.2298346      60  17:00:55

```

```
summary(m)$varcomp
```

```

          component  std.error  z.ratio bound %ch
Run          -0.004504789  0.002896281 -1.555370      U  0.1
Run:Plant!R  0.229834648  0.042316936  5.431269      P  0.0

```

Now the Run component is negative and the Run:Plant variance estimate is now equal to that for the Residuals Mean Sq for Run:Plant from the anova table.

### Set up an asrtests object

```

current.asrt <- as.asrtests(m)
print(current.asrt, which = "pseudoanova")

```

```
#### Pseudo-anova table for fixed terms
```

Wald tests for fixed effects.

Response: logitP

	Df	denDF	F.inc	Pr
(Intercept)	1	1	1550.00	0.0162
Host	1	59	59.17	0.0000
Cadavers	2	59	37.04	0.0000
Ladybird	1	59	48.26	0.0000
Host:Cadavers	2	59	0.67	0.5158
Host:Ladybird	1	59	0.99	0.3234
Cadavers:Ladybird	2	59	3.77	0.0287
Host:Cadavers:Ladybird	2	59	0.44	0.6493

The `asrtests` object contains a `wald.tab` component which can be printed by specifying that the `pseudoanova` is printed. The  $F$ -values for the fixed terms in this table are the same as those in the `anova` table.

### Obtain the marginality matrix for the fixed terms

The `pstructure` function from the `dae` package (Brien, 2024b) produce the marginality matrix for a formula as a side effect and we take advantage of that to obtain the matrix required here.

```
Ladybird.pstr <- pstructure(formula = ~ Host*Cadavers*Ladybird,  
                           data = Ladybird.dat)  
HCL.marg <- marginality(Ladybird.pstr)  
print(HCL.marg)
```

	Host	Cadavers	Host:Cadavers	Ladybird	Host:Ladybird	Cadavers:Ladybird
Host	1	0	1	0	1	0
Cadavers	0	1	1	0	0	1
Host:Cadavers	0	0	1	0	0	0
Ladybird	0	0	0	1	1	1
Host:Ladybird	0	0	0	0	1	0
Cadavers:Ladybird	0	0	0	0	0	1
Host:Cadavers:Ladybird	0	0	0	0	0	0
Host:Cadavers:Ladybird						
Host			1			
Cadavers			1			
Host:Cadavers			1			
Ladybird			1			
Host:Ladybird			1			
Cadavers:Ladybird			1			
Host:Cadavers:Ladybird			1			

This marginality matrix is interpreted by taking a row term and noting that it is marginal to any column term with a one in this row.

## Choose marginality-compliant model

```
chosen <- chooseModel(current.asrt, terms.marginality = HCL.marg)
current.asrt <- chosen$asrtests.obj
print(current.asrt, which = "test", omit.columns = c("AIC", "BIC"))
```

```
#### Sequence of model investigations for logitP
```

	terms	DF	denDF	p	action
1	Host:Cadavers:Ladybird	2	59	0.6493	Nonsignificant
2	Cadavers:Ladybird	2	59	0.0287	Significant
3	Host:Ladybird	1	59	0.3234	Nonsignificant
4	Host:Cadavers	2	59	0.5158	Nonsignificant
5	Host	1	59	0.0000	Significant

```
(chosen$sig.terms)
```

```
[[1]]
[1] "Cadavers:Ladybird"
```

```
[[2]]
[1] "Host"
```

The `chooseModel` function produces a list with components `sig.terms`, a list with the terms in the marginality-compliant model, and `asrtests.obj`, the `asrtests` object resulting from the model selection. In particular, the `asrtests` object contains a `test.summary` that details the tests performed in choosing the model; the AIC and BIC columns are omitted from `test.summary` because their inclusion has not been requested. Note that `chooseModel` does not test the main effects for Cadavers or Ladybird, because these are marginal to the significant two-factor interaction Cadavers:Ladybird.

## Form formula for selected model

```
chosen.mod <- paste(unlist(chosen$sig.terms), collapse = " + ")
(chosen.mod <- as.formula(paste("~", chosen.mod)))
```

```
~Cadavers:Ladybird + Host
```

## Obtain predictions under the chosen model and form an alldiffs object

```
diffs <- predictPlus(current.asrt$asreml.obj,
  classify = "Host:Ladybird:Cadavers",
  linear.transformation = ~Cadavers:Ladybird + Host,
  wald.tab = current.asrt$wald.tab,
  error.intervals = "halfLeast",
  LSDtype = "factor.combination", LSDby = "Host",
  tables = "predictions")
```

Joining with 'by = join\_by(fac.comb)'  
 Joining with 'by = join\_by(Host)'

#### Predictions for logitP transform(s) from Host:Ladybird:Cadavers

Notes:

- The original predictions, obtained as described below, have been linearly transformed to form estimated marginal means.
- The predictions are obtained by averaging across the hypertable calculated from model terms constructed solely from factors in the averaging and classify sets.
  - Use 'average' to move ignored factors into the averaging set.
  - The ignored set: Run

	Host	Ladybird	Cadavers	predicted.value	standard.error	upper.halfLeastSignificant.limit
1	bean	-	5	-1.6038338	0.1417454	-1.4080222
2	bean	-	10	-1.1454308	0.1417454	-0.9496192
3	bean	-	20	-0.7448097	0.1417454	-0.5489981
4	bean	+	5	-1.0195475	0.1417454	-0.8237359
5	bean	+	10	-0.5983440	0.1417454	-0.4025323
6	bean	+	20	0.4786704	0.1417454	0.6744820
7	trefoil	-	5	-2.4730339	0.1417454	-2.2772223
8	trefoil	-	10	-2.0146309	0.1417454	-1.8188193
9	trefoil	-	20	-1.6140098	0.1417454	-1.4181982
10	trefoil	+	5	-1.8887476	0.1417454	-1.6929360
11	trefoil	+	10	-1.4675441	0.1417454	-1.2717325
12	trefoil	+	20	-0.3905297	0.1417454	-0.1947181
				lower.halfLeastSignificant.limit	est.status	
1				-1.7996454	Estimable	
2				-1.3412425	Estimable	
3				-0.9406214	Estimable	
4				-1.2153592	Estimable	
5				-0.7941556	Estimable	
6				0.2828588	Estimable	
7				-2.6688455	Estimable	
8				-2.2104426	Estimable	
9				-1.8098215	Estimable	
10				-2.0845593	Estimable	
11				-1.6633557	Estimable	
12				-0.5863414	Estimable	

LSD values

minimum LSD = 0.3916233 0.3916233

mean LSD = 0.3916233 0.3916233

maximum LSD = 0.3916233 0.3916233

(sed range / mean sed = 9.92e-16 8.5e-16 )

Setting the `terms` argument to `Host:Ladybird:Cadavers` requests predictions for all combinations of the three factors and the `linear.transformation` argument is used to obtain estimated marginal means (EMMs) that conform to the chosen model. The `wald.tab` is supplied so that it can be used to get the degrees of freedom for the *t*-value to be used in calculating the LSD; the degrees of freedom to the source for the `terms` argument will be used. The `error.intervals` argument has been set to `"halfLeast"`, the `LSDtype` argument to `"factor.combination"` and the `LSDby` argument to `"Host"` so that the average LSD will be calculated for each Host. This necessary because, under the chosen model, the LSDs differ between Hosts. It results in `lower.halfLeastSignificant.limit` and `upper.halfLeastSignificant.limit` being added to the `predictions` component of the `alldiffs` object.

Or, calculate predictions to check first and then transform to conform to chosen model

```
diffs.full <- predictPlus(current.asrt$asreml.obj,
  classify = "Host:Ladybird:Cadavers",
  wald.tab = current.asrt$wald.tab,
  tables = "none", Vmatrix = TRUE)

diffs <- linTransform(diffs.full, linear.transformation = ~Cadavers:Ladybird + Host,
  wald.tab = current.asrt$wald.tab,
  error.intervals = "halfLeast",
  LSDtype = "factor.combination", LSDby = "Host",
  tables = "predictions")
```

Joining with `'by = join_by(fac.comb)'`

Joining with `'by = join_by(Host)'`

#### Predictions for logitP transform(s) from Host:Ladybird:Cadavers

Notes:

- The original predictions, obtained as described below, have been linearly transformed to form estimated marginal means.
- The predictions are obtained by averaging across the hypertable calculated from model terms constructed solely from factors in the averaging and classify sets.
  - Use `'average'` to move ignored factors into the averaging set.
  - The ignored set: Run

	Host	Ladybird	Cadavers	predicted.value	standard.error	upper.halfLeastSignificant.limit
1	bean	-	5	-1.6038338	0.1417454	-1.4080222
2	bean	-	10	-1.1454308	0.1417454	-0.9496192
3	bean	-	20	-0.7448097	0.1417454	-0.5489981
4	bean	+	5	-1.0195475	0.1417454	-0.8237359
5	bean	+	10	-0.5983440	0.1417454	-0.4025323
6	bean	+	20	0.4786704	0.1417454	0.6744820
7	trefoil	-	5	-2.4730339	0.1417454	-2.2772223
8	trefoil	-	10	-2.0146309	0.1417454	-1.8188193
9	trefoil	-	20	-1.6140098	0.1417454	-1.4181982
10	trefoil	+	5	-1.8887476	0.1417454	-1.6929360
11	trefoil	+	10	-1.4675441	0.1417454	-1.2717325

	12	trefoil	+	20	-0.3905297	0.1417454	-0.1947181
		lower.halfLeastSignificant.limit		est.status			
1		-1.7996454		Estimable			
2		-1.3412425		Estimable			
3		-0.9406214		Estimable			
4		-1.2153592		Estimable			
5		-0.7941556		Estimable			
6		0.2828588		Estimable			
7		-2.6688455		Estimable			
8		-2.2104426		Estimable			
9		-1.8098215		Estimable			
10		-2.0845593		Estimable			
11		-1.6633557		Estimable			
12		-0.5863414		Estimable			

LSD values

minimum LSD = 0.3916233 0.3916233

mean LSD = 0.3916233 0.3916233

maximum LSD = 0.3916233 0.3916233

(sed range / mean sed = 9.92e-16 8.5e-16 )

The above LSD values can only be used to compare pairs of EMMs for the same Host.

## Explore the LSDs

To investigate the errors that would result from using the overall LSDs as opposed to the LSDs computed for each Host, the `exploreLSDs`, `pickLSDstatistics` and `plotLSDerrors` functions are used, firstly with the default value of "overall" for `LSDtype` and finally with the `LSDtype` set to "factor.combination" and `LSDby` to "Host".

The `exploreLSDs` function produces tables of statistics for the LSDs computed for the settings of the `LSDtype` and `LSDby`; the settings of these arguments does not have to match those used in producing the `alldiffs` object. For `LSDtype` set to "overall", a single LSD statistic is computed that is based on the standard errors of all pairwise differences. To ascertain the errors that arise from using this LSD value for determining, for all pairwise comparisons, whether a comparison is significant, `exploreLSDs` compares the results using the LSD value with the *p*-values in the `p.differences` component of the `alldiffs` object. For `LSDtype` set to "factor.combination" and `LSDby` to "Host", the LSD statistics are calculated from standard errors of the pairwise differences for each Host. Examination of the `sed` component of `diffs` reveals that there are only three different values for the standard errors of pairwise differences and, hence, only three unique values for the LSD.

The function `pickLSDstatistics` can be used to pick a statistic that minimizes the number of false negative errors i.e. declaring a pairwise difference to be nonsignificant when it is significant. The function has an argument `false.pos.wt` that specifies how many false negatives are equivalent to one false positive, a false positive occurring when a pairwise difference that is nonsignificant is declared to be significant; it allows the choice of an LSD statistic that balances the number of false positives and negatives. Here are the tables of the numbers of false positive and negative error in using the values of the various LSD statistics for determining the significance of the 66 pairwise comparisons of the 12 predictions.



```
exploreLSDs(diffs, LSDtype = "overall")
```

```
#### Statistics calculated from LSD values
```

	c	min	quant10	quant25	mean	median	quant75	quant90	max
1	66	0.2261038	0.3916233	0.3916233	0.4090372	0.3916233	0.4522076	0.4522076	0.4522076

```
#### False positives resulting from the use of various LSD statistics
```

	c	min	quant10	quant25	mean	median	quant75	quant90	max
false.pos	66	7	2	2	2	2	0	0	0

```
#### False negatives resulting from the use of various LSD statistics
```

	c	min	quant10	quant25	mean	median	quant75	quant90	max
false.neg	66	0	0	0	2	0	4	4	4

```
(pickLSDstatistics(diffs))
```

```
[1] "q75"
```

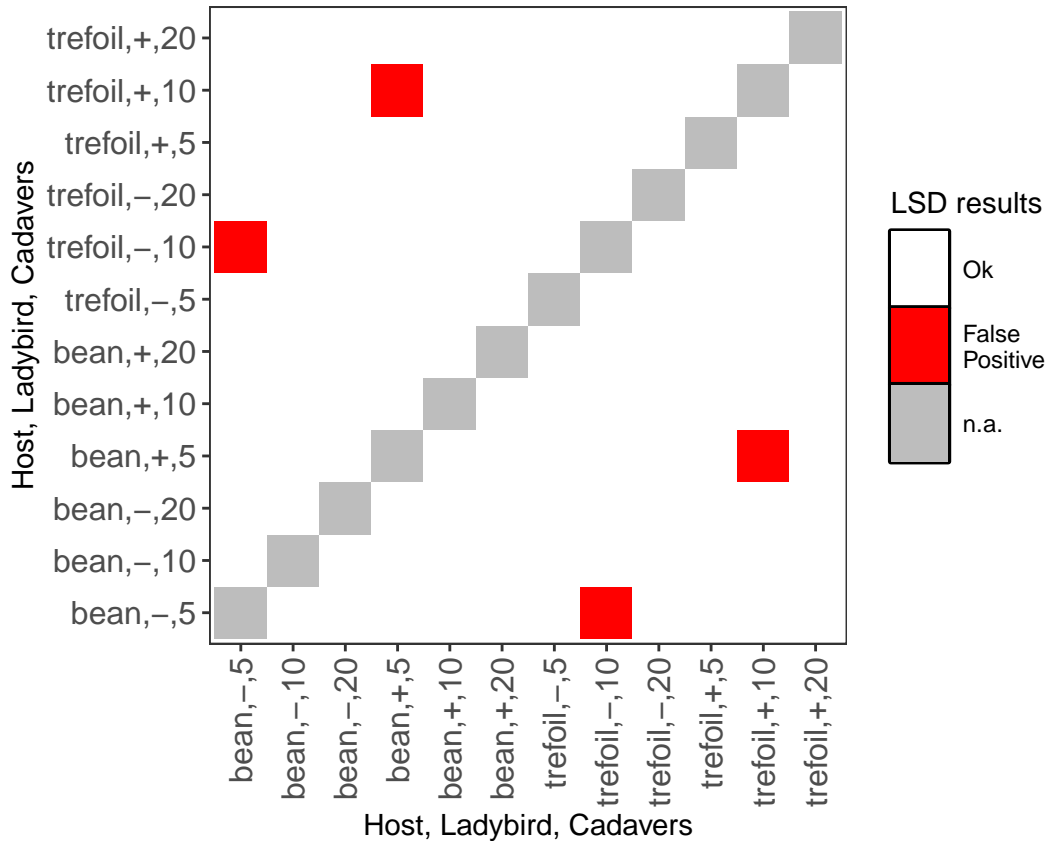
```
(pickLSDstatistics(diffs, false.pos.wt = 1))
```

```
[1] "q10"
```

Of the statistics shown, there is no choice that will not result in errors. Using the 75th quantile of the all LSD values for all pairwise comparisons (`quant75`) will result in minimum number of false negatives, without there being any false positives; in this case, there will be four out of the 66 comparisons whose conclusions will be false negatives. If one is prepared to consider false negatives and positives as being equally bad, then set `false.pos.wt` to one and the 10th quantile of the LSD values (`quant10`) will be chosen as the smallest value of the LSD statistics that has the minimum weighted sum of the errors; using it will result in two of the 66 comparisons yielding false positive results.

To see which of the two pairwise comparisons will be falsely identified as being significant when the LSD value is set to `quant10`, the LSDs stored in the `alldiffs` object need to be recalculated to be based on the value for this statistic. Then plot the errors or save the return values obtained using the function `plotLSDerrors`. The plot below shows that the two pairs whose differences are incorrectly identified as significant have the same level of the Ladybird factor, but differ in both of the levels for the Host and Cadaver factors.

```
diffs.overall <- recalcLSD(diffs, LSDtype = "overall", LSDstatistic = "q10")
plotLSDerrors(diffs.overall)
```



This raises the question of whether the 10th quantile of all of the LSDs should be used. There are at least four alternatives: (i) use it without restriction, on the basis that it can be concluded that using it is unlikely to result in seriously flawed conclusions; (ii) use it with the restriction that it only be applied to assess pairwise comparisons that have the same Host or the same Cadaver treatment; (iii) investigate the use of an overall LSD based on `quant75`; and (iv) rather than use an overall LSD value, use LSD values computed from the LSDs within each Host level.

Because `LSDtype` was set to `"factor.combination"` and `LSDby` to `"Host"` in forming the object `diffs`, the LSDs for alternative (iv) are stored in the `LSD` component of the object `diffs`. Printing out the `LSD` component will summarize how those LSD values perform. Otherwise, the following call to `exploreLSDs` will display the properties of the LSDs for various LSD statistics:

```
exploreLSDs(diffs, LSDtype = "factor.combination", LSDby = "Host")
```

The following shows the contents of the `LSD` component of `diffs`:

```
(diffs$LSD)
      c  minLSD  meanLSD  maxLSD assignedLSD  accuracyLSD  falsePos  falseNeg
bean  15 0.3916233 0.3916233 0.3916233  0.3916233 5.669852e-16      0      0
trefoil 15 0.3916233 0.3916233 0.3916233  0.3916233 5.669852e-16      0      0
```

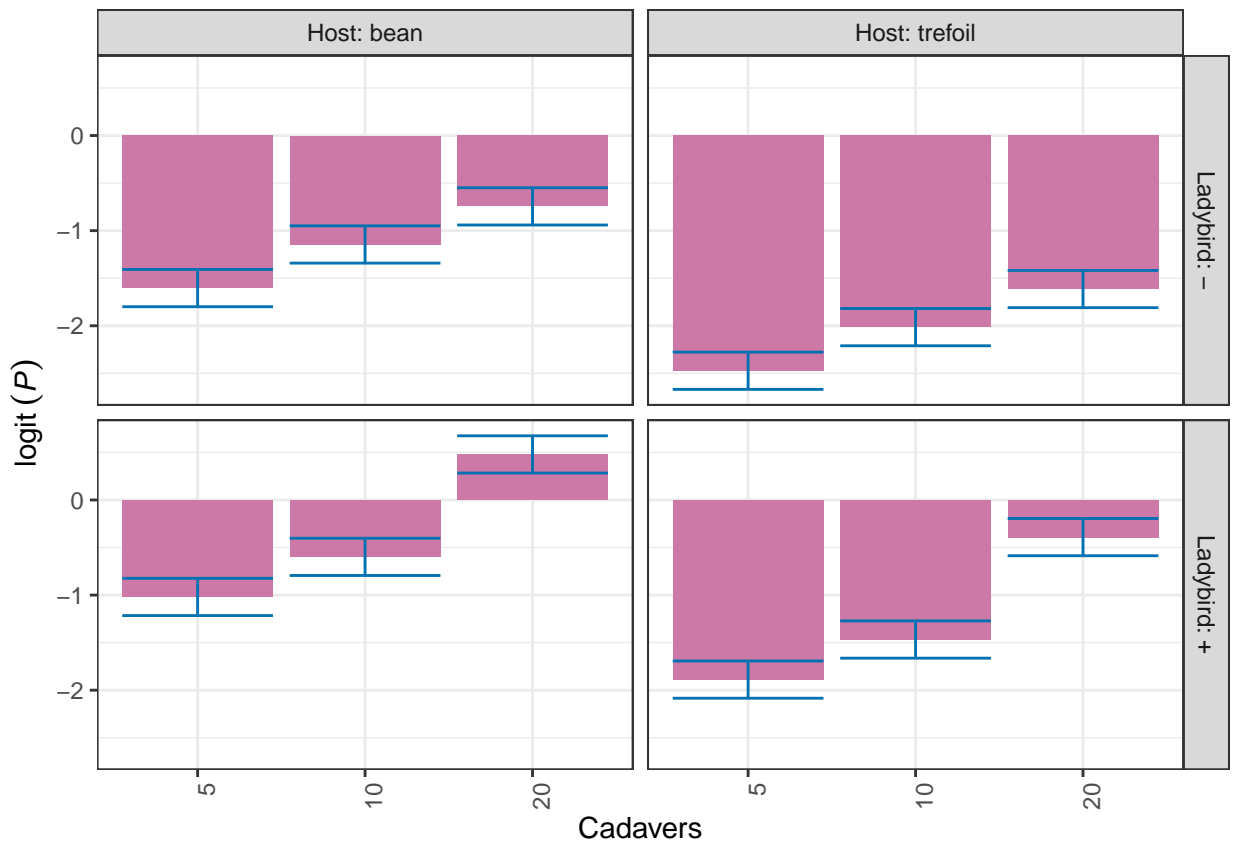
Because the minimum and maximum LSD values are equal, it follows that there is only one value of the LSD for all pairwise comparisons within each Host level and it happens that the values for the two Hosts are also equal. The table shows that zero false positives and negatives will result from the use of the value of 0.39 for the 30 within-Host comparisons.

## Plot the predictions

```

titl <- str2expression("logit~(italic(P))")
names(titl) <- "logitP"
plotPredictions(diffs$predictions, y = "predicted.value",
  y.title = titl,
  classify = "Host:Ladybird:Cadavers",
  error.intervals = "halfLeast", interval.annotate = FALSE,
  ggplotFuncs = list(facet_grid(Ladybird ~ Host,
    labeller = label_both)))

```



**Figure 1.** Estimated marginal means (EMMs) for  $\text{logit}(P)$ , where  $P$  is the proportion of live aphids that were infected, for two Hosts, two Ladybird levels and three Cadaver levels. Error bars are an EMM  $\pm$  half-LSD (5%). The two EMMs for the same Host are significantly different ( $p \leq 0.05$ ) if their error bars do not overlap.

The function `plotPredictions` uses `ggplot` to produce the plot and the `ggplotFuncs` argument allows the addition of `ggplot` functions to modify the plot. In this case, the `facet.grid` function is respescribed to include `prepender` functions that modify the labels of the facets to include the factor names. Note the statement in the legend of Figure 1 that restricts the use of the error bars to determining the significance of differences for the pairwise comparisons of EMMs for the same Host.

## Get and plot the predictions with a single function call

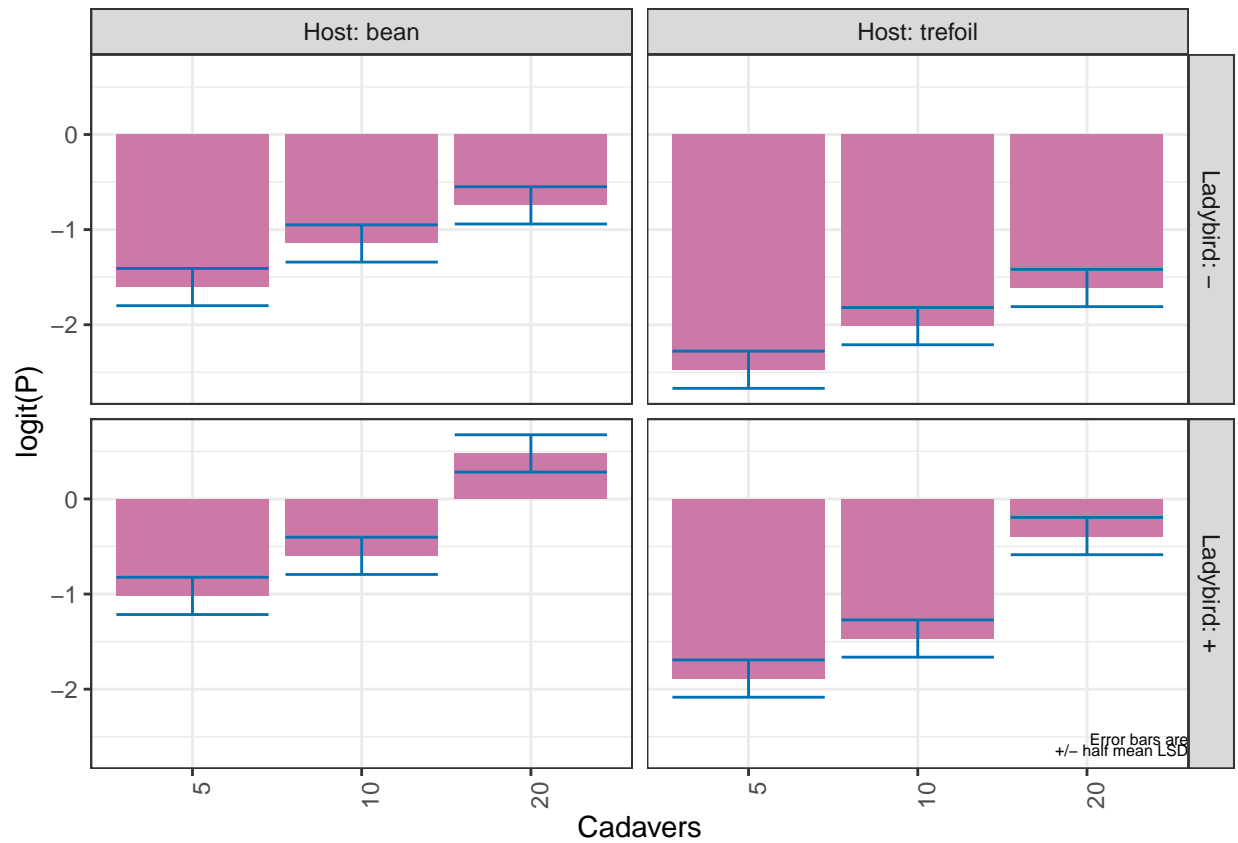
The `predictPresent` function combines the functionality of `predictPlus` and `plotPredictions`, as demonstrated now. Also, the use of `plotPvalues` to plot the pairwise  $p$ -values is displayed. The `predictPresent` function has the capability of producing `alldiffs` objects for multiple terms and these are stored in a list each of which is named for the term whose `alldiffs` object it stores. Thus, the term has to be specified in referencing components of `diffs`.

```
titl <- "logit(P)"
names(titl) <- "logitP"
diffs <- predictPresent(current.asrt$asreml.obj,
  terms = "Host:Ladybird:Cadavers",
  linear.transformation = ~Cadavers:Ladybird + Host,
  titles = titl,
  wald.tab = current.asrt$wald.tab,
  error.intervals = "halfLeast",
  LSDtype = "factor.combination", LSDby = "Host",
  tables = "none",
  ggplotFuncs = list(facet_grid(Ladybird ~ Host,
    labeller = label_both)))
```

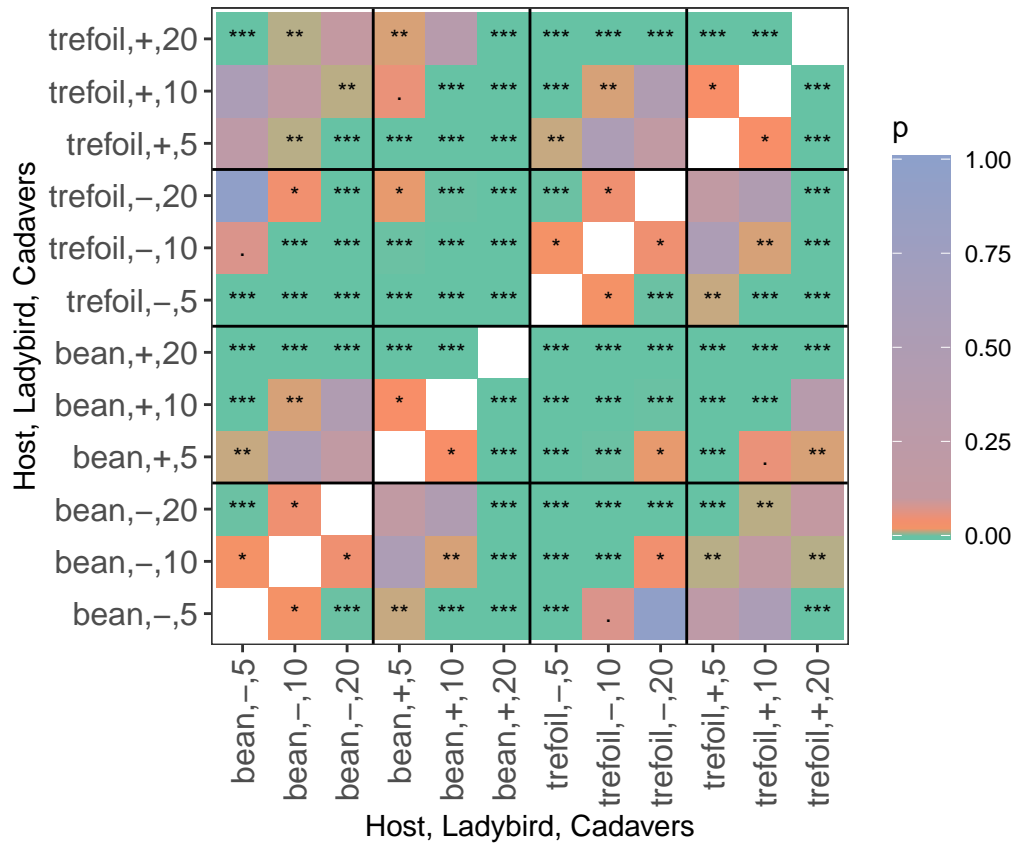
Joining with 'by = join\_by(fac.comb)'

Joining with 'by = join\_by(Host)'

```
plotPvalues(diffs$Host.Ladybird.Cadavers, factors.per.grid = 1, show.sig = TRUE)
```



**Figure 2.** Estimated marginal means (EMMs) for  $\text{logit}(P)$ , where  $P$  is the proportion of live aphids that were infected, for two Hosts, two Ladybird levels and three Cadaver levels. Error bars are an EMM  $\pm$  half-LSD (5%). The two EMMs for the same Host are significantly different ( $p \leq 0.05$ ) if their error bars do not overlap.



**Figure 3.** The  $p$ -values for each of the pairwise comparisons of the estimated marginal means for  $\text{logit}(P)$ , where  $P$  is the proportion of live aphids that were infected, for two Hosts, two Ladybird levels and three Cadaver levels

```
options(width = 90)
diffs$Host.Ladybird.Cadavers$differences
```

	bean,-,5	bean,-,10	bean,-,20	bean,+,5	bean,+,10	bean,+,20
bean,-,5	0.0000000	-0.4584030	-0.8590241	-0.5842863	-1.0054898	-2.0825042
bean,-,10	0.45840297	0.0000000	-0.4006211	-0.1258833	-0.5470869	-1.6241012
bean,-,20	0.85902408	0.4006211	0.0000000	0.2747378	-0.1464657	-1.2234801
bean,+,5	0.58428627	0.1258833	-0.2747378	0.0000000	-0.4212036	-1.4982179
bean,+,10	1.00548982	0.5470869	0.1464657	0.4212036	0.0000000	-1.0770144
bean,+,20	2.08250420	1.6241012	1.2234801	1.4982179	1.0770144	0.0000000
trefoil,-,5	-0.86920012	-1.3276031	-1.7282242	-1.4534864	-1.8746899	-2.9517043
trefoil,-,10	-0.41079715	-0.8692001	-1.2698212	-0.9950834	-1.4162870	-2.4933014
trefoil,-,20	-0.01017604	-0.4685790	-0.8692001	-0.5944623	-1.0156659	-2.0926802
trefoil,+,5	-0.28491385	-0.7433168	-1.1439379	-0.8692001	-1.2904037	-2.3674180
trefoil,+,10	0.13628970	-0.3221133	-0.7227344	-0.4479966	-0.8692001	-1.9462145
trefoil,+,20	1.21330408	0.7549011	0.3542800	0.6290178	0.2078143	-0.8692001
	trefoil,-,5	trefoil,-,10	trefoil,-,20	trefoil,+,5	trefoil,+,10	trefoil,+,20
bean,-,5	0.8692001	0.4107972	0.01017604	0.2849139	-0.1362897	-1.2133041
bean,-,10	1.3276031	0.8692001	0.46857901	0.7433168	0.3221133	-0.7549011
bean,-,20	1.7282242	1.2698212	0.86920012	1.1439379	0.7227344	-0.3542800
bean,+,5	1.4534864	0.9950834	0.59446231	0.8692001	0.4479966	-0.6290178
bean,+,10	1.8746899	1.4162870	1.01566586	1.2904037	0.8692001	-0.2078143
bean,+,20	2.9517043	2.4933014	2.09268024	2.3674180	1.9462145	0.8692001
trefoil,-,5	0.0000000	-0.4584030	-0.85902408	-0.5842863	-1.0054898	-2.0825042
trefoil,-,10	0.4584030	0.0000000	-0.40062111	-0.1258833	-0.5470869	-1.6241012
trefoil,-,20	0.8590241	0.4006211	0.00000000	0.2747378	-0.1464657	-1.2234801
trefoil,+,5	0.5842863	0.1258833	-0.27473781	0.0000000	-0.4212036	-1.4982179
trefoil,+,10	1.0054898	0.5470869	0.14646574	0.4212036	0.0000000	-1.0770144
trefoil,+,20	2.0825042	1.6241012	1.22348012	1.4982179	1.0770144	0.0000000

```
options(width = 90)
print(diffs$Host.Ladybird.Cadavers$sed)
```

	bean,-,5	bean,-,10	bean,-,20	bean+,5	bean+,10	bean+,20	trefoil,-,5
bean,-,5	NA	0.1957142	0.1957142	0.1957142	0.1957142	0.1957142	0.1129957
bean,-,10	0.1957142	NA	0.1957142	0.1957142	0.1957142	0.1957142	0.2259913
bean,-,20	0.1957142	0.1957142	NA	0.1957142	0.1957142	0.1957142	0.2259913
bean+,5	0.1957142	0.1957142	0.1957142	NA	0.1957142	0.1957142	0.2259913
bean+,10	0.1957142	0.1957142	0.1957142	0.1957142	NA	0.1957142	0.2259913
bean+,20	0.1957142	0.1957142	0.1957142	0.1957142	0.1957142	NA	0.2259913
trefoil,-,5	0.1129957	0.2259913	0.2259913	0.2259913	0.2259913	0.2259913	NA
trefoil,-,10	0.2259913	0.1129957	0.2259913	0.2259913	0.2259913	0.2259913	0.1957142
trefoil,-,20	0.2259913	0.2259913	0.1129957	0.2259913	0.2259913	0.2259913	0.1957142
trefoil+,5	0.2259913	0.2259913	0.2259913	0.1129957	0.2259913	0.2259913	0.1957142
trefoil+,10	0.2259913	0.2259913	0.2259913	0.2259913	0.1129957	0.2259913	0.1957142
trefoil+,20	0.2259913	0.2259913	0.2259913	0.2259913	0.2259913	0.1129957	0.1957142
	trefoil,-,10	trefoil,-,20	trefoil+,5	trefoil+,10	trefoil+,20		
bean,-,5	0.2259913	0.2259913	0.2259913	0.2259913	0.2259913		
bean,-,10	0.1129957	0.2259913	0.2259913	0.2259913	0.2259913		
bean,-,20	0.2259913	0.1129957	0.2259913	0.2259913	0.2259913		
bean+,5	0.2259913	0.2259913	0.1129957	0.2259913	0.2259913		
bean+,10	0.2259913	0.2259913	0.2259913	0.1129957	0.2259913		
bean+,20	0.2259913	0.2259913	0.2259913	0.2259913	0.1129957		
trefoil,-,5	0.1957142	0.1957142	0.1957142	0.1957142	0.1957142		
trefoil,-,10	NA	0.1957142	0.1957142	0.1957142	0.1957142		
trefoil,-,20	0.1957142	NA	0.1957142	0.1957142	0.1957142		
trefoil+,5	0.1957142	0.1957142	NA	0.1957142	0.1957142		
trefoil+,10	0.1957142	0.1957142	0.1957142	NA	0.1957142		
trefoil+,20	0.1957142	0.1957142	0.1957142	0.1957142	NA		



## Perform the analysis with just selected model fitted

The model with nonsignificant fixed terms dropped is obtained in order to compare it with the fit when they are retained and the estimated marginal means for the chosen model are obtained.

```
ns.terms <- current.asrt$test.summary$terms[current.asrt$test.summary$action == "Nonsignificant"]
red.asrt <- changeTerms(current.asrt, dropFixed = paste(ns.terms, collapse = "+"))
summary(red.asrt$asreml.obj)$varcomp
```

```
          component  std.error  z.ratio bound %ch
Run          -0.004327122 0.002802876 -1.543815    U 0.8
Run:Plant!R  0.223431497 0.039503516  5.655990    P 0.0
```

```
print(red.asrt, which = "pseudoanova")
```

```
#### Pseudo-anova table for fixed terms
```

Wald tests for fixed effects.

Response: logitP

	Df	denDF	F.inc	Pr
(Intercept)	1	1	1550.00	0.0162
Host	1	64	60.88	0.0000
Cadavers	2	64	38.12	0.0000
Ladybird	1	64	49.65	0.0000
Cadavers:Ladybird	2	64	3.88	0.0256

```
diffs.red <- predictPlus(red.asrt$asreml.obj,
  classify = "Host:Ladybird:Cadavers",
  wald.tab = current.asrt$wald.tab,
  error.intervals = "halfLeast", interval.annotate = FALSE,
  LSDtype = "factor.combination", LSDby = "Host",
  tables = "predictions")
```

```
Joining with 'by = join_by(fac.comb)'
```

```
Joining with 'by = join_by(Host)'
```

```
#### Predictions for logitP from Host:Ladybird:Cadavers
```

Notes:

- The predictions are obtained by averaging across the hypertable calculated from model terms constructed solely from factors in the averaging and classify sets.
- Use 'average' to move ignored factors into the averaging set.
- The ignored set: Run

	Host	Ladybird	Cadavers	predicted.value	standard.error
1	bean	-	5	-1.6038338	0.1398332
2	bean	-	10	-1.1454308	0.1398332
3	bean	-	20	-0.7448097	0.1398332
4	bean	+	5	-1.0195475	0.1398332
5	bean	+	10	-0.5983440	0.1398332
6	bean	+	20	0.4786704	0.1398332
7	trefoil	-	5	-2.4730339	0.1398332
8	trefoil	-	10	-2.0146309	0.1398332
9	trefoil	-	20	-1.6140098	0.1398332
10	trefoil	+	5	-1.8887476	0.1398332
11	trefoil	+	10	-1.4675441	0.1398332
12	trefoil	+	20	-0.3905297	0.1398332

	upper.halfLeastSignificant.limit	lower.halfLeastSignificant.limit	est.status
1	-1.4107942	-1.7968734	Estimable
2	-0.9523913	-1.3384704	Estimable
3	-0.5517702	-0.9378493	Estimable
4	-0.8265080	-1.2125871	Estimable
5	-0.4053044	-0.7913835	Estimable
6	0.6717100	0.2856308	Estimable
7	-2.2799944	-2.6660735	Estimable
8	-1.8215914	-2.2076705	Estimable
9	-1.4209703	-1.8070494	Estimable
10	-1.6957081	-2.0817872	Estimable
11	-1.2745045	-1.6605837	Estimable
12	-0.1974902	-0.5835693	Estimable

LSD values

minimum LSD = 0.3860791 0.3860791

mean LSD = 0.3860791 0.3860791

maximum LSD = 0.3860791 0.3860791

(sed range / mean sed = 1.44e-16 1.44e-16 )

```
options(width = 90)
print(diffs.red$sed)
```

	bean,-,5	bean,-,10	bean,-,20	bean,+,5	bean,+,10	bean,+,20	trefoil,-,5
bean,-,5	NA	0.1929435	0.1929435	0.1929435	0.1929435	0.1929435	0.1113960
bean,-,10	0.1929435	NA	0.1929435	0.1929435	0.1929435	0.1929435	0.2227920
bean,-,20	0.1929435	0.1929435	NA	0.1929435	0.1929435	0.1929435	0.2227920
bean,+,5	0.1929435	0.1929435	0.1929435	NA	0.1929435	0.1929435	0.2227920
bean,+,10	0.1929435	0.1929435	0.1929435	0.1929435	NA	0.1929435	0.2227920
bean,+,20	0.1929435	0.1929435	0.1929435	0.1929435	0.1929435	NA	0.2227920
trefoil,-,5	0.1113960	0.2227920	0.2227920	0.2227920	0.2227920	0.2227920	NA
trefoil,-,10	0.2227920	0.1113960	0.2227920	0.2227920	0.2227920	0.2227920	0.1929435
trefoil,-,20	0.2227920	0.2227920	0.1113960	0.2227920	0.2227920	0.2227920	0.1929435
trefoil,+,5	0.2227920	0.2227920	0.2227920	0.1113960	0.2227920	0.2227920	0.1929435
trefoil,+,10	0.2227920	0.2227920	0.2227920	0.2227920	0.1113960	0.2227920	0.1929435

trefoil,+,20	0.2227920	0.2227920	0.2227920	0.2227920	0.2227920	0.1113960	0.1929435
	trefoil,-,10	trefoil,-,20	trefoil,+,5	trefoil,+,10	trefoil,+,20		
bean,-,5	0.2227920	0.2227920	0.2227920	0.2227920	0.2227920	0.2227920	
bean,-,10	0.1113960	0.2227920	0.2227920	0.2227920	0.2227920	0.2227920	
bean,-,20	0.2227920	0.1113960	0.2227920	0.2227920	0.2227920	0.2227920	
bean,+,5	0.2227920	0.2227920	0.1113960	0.2227920	0.2227920	0.2227920	
bean,+,10	0.2227920	0.2227920	0.2227920	0.1113960	0.2227920	0.2227920	
bean,+,20	0.2227920	0.2227920	0.2227920	0.2227920	0.2227920	0.1113960	
trefoil,-,5	0.1929435	0.1929435	0.1929435	0.1929435	0.1929435	0.1929435	
trefoil,-,10	NA	0.1929435	0.1929435	0.1929435	0.1929435	0.1929435	
trefoil,-,20	0.1929435	NA	0.1929435	0.1929435	0.1929435	0.1929435	
trefoil,+,5	0.1929435	0.1929435	NA	0.1929435	0.1929435	0.1929435	
trefoil,+,10	0.1929435	0.1929435	0.1929435	NA	0.1929435	0.1929435	
trefoil,+,20	0.1929435	0.1929435	0.1929435	0.1929435	NA	0.1929435	

## References

- Brien, C. J. (2024a) `asremlPlus`: *Augments ASReml-R in fitting mixed models and packages generally in exploring prediction differences*. Version 4.4.43. <https://cran.r-project.org/package=asremlPlus/> or <http://chris.brien.name/rpackages/>.
- Brien, C. J. (2024b) `dae`: *Functions useful in the design and ANOVA of experiments*. Version 3.2.30. <https://cran.r-project.org/package=dae/> or <http://chris.brien.name/rpackages/>.
- Butler, D. G., Cullis, B. R., Gilmour, A. R., Gogel, B. J. and Thompson, R. (2023). *ASReml-R Reference Manual Version 4.2*. VSN International Ltd, <https://https://asreml.kb.vsnr.co.uk/>.
- Littell, R. C., Milliken, G. A., Stroup, W. W., Wolfinger, R. D., & Schabenberger, O. (2006). *SAS for Mixed Model.* (2nd ed.). Cary, N.C.: SAS Press.
- R Core Team (2024) *R: A language and environment for statistical computing*. Vienna, Austria: R Foundation for Statistical Computing. <https://www.r-project.org/>.
- Snee, R. D. (1981). Graphical Display and Assessment of Means. *Biometrics*, **37**, 835–836.
- Welham, S. J., Gezan, S. A., Clark, S. J., & Mead, A. (2014). *Statistical Methods in Biology: Design and Analysis of Experiments and Regression*. Boca Raton: Chapman and Hall/CRC.