

Package ‘gsean’

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Type Package

Title Gene Set Enrichment Analysis with Networks

Description Biological molecules in a living organism seldom work individually. They usually interact each other in a cooperative way. Biological process is too complicated to understand without considering such interactions. Thus, network-based procedures can be seen as powerful methods for studying complex process. However, many methods are devised for analyzing individual genes. It is said that techniques based on biological networks such as gene co-expression are more precise ways to represent information than those using lists of genes only. This package is aimed to integrate the gene expression and biological network. A biological network is constructed from gene expression data and it is used for Gene Set Enrichment Analysis.

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R topics documented:

gsean-package	2
centrality_gsea	3
exprs2adj	4
GO_dme	5
gsean	5
KEGG_hsa	7
label_prop_gsea	7

Index	9
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gsean-package

Gene Set Enrichment Analysis with Networks

Description

Biological molecules in a living organism seldom work individually. They usually interact each other in a cooperative way. Biological process is too complicated to understand without considering such interactions. Thus, network-based procedures can be seen as powerful methods for studying complex process. However, many methods are devised for analyzing individual genes. It is said that techniques based on biological networks such as gene co-expression are more precise ways to represent information than those using lists of genes only. This package is aimed to integrate the gene expression and biological network. A biological network is constructed from gene expression data and it is used for Gene Set Enrichment Analysis.

Details

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Index: This package was not yet installed at build time.

Author(s)

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`centrality_gsea`*Gene Set Enrichment Analysis with centrality measure*

Description

GSEA is performed with centrality measure

Usage

```
centrality_gsea(geneset, x, adjacency, pseudo = 1, nperm = 1000,  
               centrality = function(x) rowSums(abs(x)),  
               weightParam = 1, minSize = 1, maxSize = Inf,  
               gseaParam = 1, nproc = 0, BPPARAM = NULL)
```

Arguments

<code>geneset</code>	list of gene sets
<code>x</code>	Named vector of gene-level statistics. Names should be the same as in gene sets.
<code>adjacency</code>	adjacency matrix
<code>pseudo</code>	pseudo number for log2 transformation (default: 1)
<code>nperm</code>	number of permutations (default: 1000)
<code>centrality</code>	centrality measure, degree centrality or node strength is default
<code>weightParam</code>	weight parameter value for the centrality measure, equally weight if weightParam = 0 (default: 1)
<code>minSize</code>	minimal size of a gene set (default: 1)
<code>maxSize</code>	maximal size of a gene set (default: Inf)
<code>gseaParam</code>	GSEA parameter value (default: 1)
<code>nproc</code>	see <code>fgsea::fgsea</code>
<code>BPPARAM</code>	see <code>fgsea::fgsea</code>

Value

GSEA result

Author(s)

Dongmin Jung

See Also

`fgsea::fgsea`

Examples

```
data(examplePathways)
data(exampleRanks)
exampleRanks <- exampleRanks[1:100]
adjacency <- diag(length(exampleRanks))
rownames(adjacency) <- names(exampleRanks)
set.seed(1)
result.GSEA <- centrality_gsea(examplePathways, exampleRanks, adjacency)
```

exprs2adj	<i>Convert gene expression data to adjacency matrix by using correlation coefficients</i>
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Description

A biological network is constructed from gene expression data and it is used for Gene Set Enrichment Analysis.

Usage

```
exprs2adj(x, pseudo = 1, ...)
```

Arguments

x	gene expression data
pseudo	pseudo number for log2 transformation (default: 1)
...	additional parameters for correlation; see WGCNA::cor

Value

adjacency matrix

Author(s)

Dongmin Jung

See Also

fgsea::fgsea, WGCNA::cor

Examples

```
data(exampleRanks)
Names <- names(exampleRanks)
exprs <- matrix(rnorm(10*length(exampleRanks)), ncol = 10)
adjacency <- exprs2adj(exprs)
```

GO_dme

Gene Ontology terms with gene ID for Drosophila melanogaster

Description

The data set contains all Gene Ontology terms for Drosophila melanogaster and genes are identified by gene ID. There are 2823 categories.

Usage

GO_dme

Format

a list of gene sets

Value

GO gene sets

Author(s)

Dongmin Jung

Source

<http://www.go2msig.org/cgi-bin/prebuilt.cgi?taxid=7227>

Examples

```
load(system.file("data", "GO_dme.rda", package = "gsean"))
```

gsean

Gene Set Enrichment Analysis with Networks

Description

GSEA or ORA is performed with networks from gene expression data

Usage

```
gsean(geneset, x, exprs, pseudo = 1, threshold = 0.99, nperm = 1000,  
      centrality = function(x) rowSums(abs(x)), weightParam = 1,  
      minSize = 1, maxSize = Inf, gseaParam = 1, nproc = 0,  
      BPPARAM = NULL, corParam = list(), tmax = 10, ...)
```

Arguments

geneset	list of gene sets
x	Named vector of gene-level statistics for GSEA or set of genes for ORA. Names should be the same as in gene sets.
exprs	gene expression data
pseudo	pseudo number for log2 transformation (default: 1)
threshold	threshold of correlation for nodes to be considered neighbors for ORA (default: 0.99)
nperm	number of permutations (default: 1000)
centrality	centrality measure, degree centrality or node strength is default
weightParam	weight parameter value for the centrality measure, equally weight if weightParam = 0 (default: 1)
minSize	minimal size of a gene set (default: 1)
maxSize	maximal size of a gene set (default: Inf)
gseaParam	GSEA parameter value (default: 1)
nproc	see fgsea::fgsea
BPPARAM	see fgsea::fgsea
corParam	additional parameters for correlation; see WGCNA::cor
tmax	maximum number of iterations for label propagation (default: 10)
...	additional parameters for label propagation; see RANKS::label.prop

Value

GSEA result

Author(s)

Dongmin Jung

See Also

exprs2adj, label_prop_gsea, centrality_gsea

Examples

```
data(examplePathways)
data(exampleRanks)
exampleRanks <- exampleRanks[1:100]
Names <- names(exampleRanks)
exprs <- matrix(rnorm(10*length(exampleRanks)), ncol = 10)
rownames(exprs) <- names(exampleRanks)
set.seed(1)
result.GSEA <- gsean(examplePathways, exampleRanks, exprs)
```

KEGG_hsa

KEGG pathways with gene symbol for human

Description

The data set contains 186 KEGG pathways for *Drosophila melanogaster* and genes are identified by gene symbol.

Usage

KEGG_hsa

Format

a list of gene sets

Value

KEGG gene sets

Author(s)

Dongmin Jung

Source

<http://software.broadinstitute.org/gsea/msigdb/collections.jsp>

Examples

```
load(system.file("data", "KEGG_hsa.rda", package = "gsean"))
```

label_prop_gsea

Over-representaion analysis with the label propagation algorithm

Description

ORA is performed by GSEA with the label propagation algorithm

Usage

```
label_prop_gsea(geneset, x, adjacency, threshold = 0.99, nperm = 1000,  
                minSize = 1, maxSize = Inf, gseaParam = 1, nproc = 0,  
                BPPARAM = NULL, ...)
```

Arguments

geneset	list of gene sets
x	set of genes
adjacency	adjacency matrix
threshold	threshold of correlation for nodes to be considered neighbors (default: 0.99)
nperm	number of permutations (default: 1000)
minSize	minimal size of a gene set (default: 1)
maxSize	maximal size of a gene set (default: Inf)
gseaParam	GSEA parameter value (default: 1)
nproc	see fgsea::fgsea
BPPARAM	see fgsea::fgsea
...	additional parameters for label propagation; see RANKS::label.prop

Value

GSEA result

Author(s)

Dongmin Jung

See Also

fgsea::fgsea, RANKS::label.prop

Examples

```
data(examplePathways)
data(exampleRanks)
exampleRanks <- exampleRanks[1:100]
geneNames <- names(exampleRanks)
set.seed(1)
x <- sample(geneNames, 10)
adjacency <- diag(length(exampleRanks))
rownames(adjacency) <- geneNames
result.GSEA <- label_prop_gsea(examplePathways, x, adjacency)
```


Index

centrality_gsea, [3](#)

exprs2adj, [4](#)

GO_dme, [5](#)

gsean, [5](#)

gsean-package, [2](#)

KEGG_hsa, [7](#)

label_prop_gsea, [7](#)